

# Challenges and Pragmatic Solutions to Statistical Analysis of High-throughput Genomic Data

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## Abstract

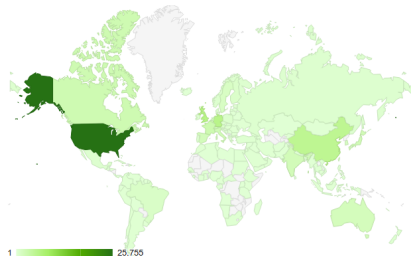
The *R / Bioconductor* project<sup>1</sup> provides a proving ground for computational approaches to handling high-volume genomic data. Many investigators have primary interests and talent in domains other than computer science. Their research questions raise transient analytic needs that make it difficult to justify narrowly-focused investment in sophisticated computational methods or machinery. Very diverse computational environments make many solutions idiosyncratic. This leads us toward development of reusable infrastructure to support simple and standardized models of high-throughput computation, relying on opportunistic community standards, and offering consistently-configured computational environments for scalable evaluation.

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<sup>1</sup><http://bioconductor.org>

# R / Bioconductor

- ▶ 610 packages for analysis and comprehension of high-throughput genomic data
- ▶ Statisticians, biologists, bioinformaticians in US, Europe, Asia, ...; mid-sized labs & researchers in academia, government, pharma
- ▶ Developed by advanced users, domain experts, core group



Google analytics, 1-month access, 10 December 2012

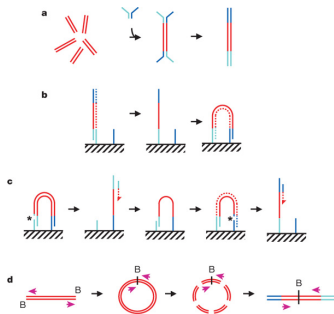
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Maintainers	Packages
340	1
68	2
16	3
9	4
3	5
5	6
1	7
1	20
1	44

# R / Bioconductor

## High-throughput genomic data

- ▶ Sequences: very large data summarized or filtered to modest size for advanced statistical analysis, e.g., *edgeR*, *DESeq*, *VariantTools*
- ▶ Variants: statistical association with phenotype; e.g., millions of SNPs  $\times$  thousands of individuals, SNPs perhaps in combination, e.g., *snpStats*, *MatrixEQTL*
- ▶ Arrays: whole-genome scans with locally complex structure, e.g., *bumphunter*



Bentley et al., Nature  
2008 456(7218):53-9

# R / Bioconductor

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## Differential expression

1. Align: third-party  $\Rightarrow$  BAM files
2. Count: 'annotation', *GenomicRanges*  
*findOverlaps*; data reduction
3. Test: microarray-like

$$\log \mu_{gi} = \mathbf{x}_i^T \beta_g + \log N_i$$

Neg. binomial GLM

Shared info. across experiment

# R / Bioconductor

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Method	500k SNPs
Plink	583.3 days
Merlin	20.0 days
R/qtl	4.7 days
<i>snpMatrix</i>	5.1 days
Matrix eQTL	19.4 mins

## Anecdote (old)

- ▶ glm, 100's / minute
- ▶ glm.fit & tricks, 1000's / minute
- ▶ Cluster: 500,000 / minute

# R / Bioconductor

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## Bump-hunting

$$Y_{ij} = \beta_0(l_j) + \beta_1(l_j)X_j + \varepsilon_{ij}$$

Subject  $i$ , location  $l_j$ , covariate  $X_j$ ; baseline function  $\beta_0(l_j)$ , parameter of interest  $\beta_1(l_j)$

Shared info. between nearby locations



# Pragmatic approaches to big data

What is needed for big data analysis?

- ▶ Efficient, robust code
- ▶ Memory management
- ▶ Parallel evaluation
- ▶ Algorithms

# Efficient, robust code

Experienced *R* programmers...

- ▶ Vectors, vs. element-wise iteration
  - ▶ `for` tempts users
- ▶ Pre-allocate & fill, vs. copy & append
  - ▶ `lapply` guides users
- ▶ Selective input
- ▶ Surprising gotchas, e.g., `unlist`  
`use.names=TRUE`, vs. `use.names=FALSE`
- ▶ Specialized packages & functions

*Anecdotal*  
(*Bioconductor*  
submission, R-help,  
StackOverflow<sup>2</sup>,  
...): a common  
shortcoming

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<sup>2</sup><http://stackoverflow.com>

# Efficient, robust code

## A common approach

- ▶ C code – directly or via add-ons like *Rcpp*

## Robust

- ▶ Developers seem to want their code to work

	Used by
src directory	232
<i>Rcpp</i>	10
<i>RUnit</i>	78
<i>testthat</i>	7

# Memory management

	Used by
SQL	43
<i>ff</i> , <i>bigmemory</i>	11
<i>ncdf</i>	3

- ▶ SQL used (appropriately) for relational data
- ▶ *R*-specific solutions require dedicated development without data re-use in other applications
- ▶ NetCDF (a standard) not widely used
  - ▶ 3rd party dependency
  - ▶ Experience of developers
  - ▶ Limitations of *R* interface

# Memory management

	Used by
SQL	43
<i>ff</i> , <i>bigmemory</i>	11
<i>ncdf</i>	3

- ▶ Large vectors probably do not play well with using multiple cores (though what is large?)
- ▶ Instead: data slices, iteration, on-line algorithms; data containers, e.g., *IRanges::Rle*-class

# Parallel evaluation

	Used by
<i>parallel</i>	26
<i>snow</i> & c.	20
<i>foreach</i> & c.	11
<i>rlecuyer</i> , <i>setRNG</i>	2
<i>Rmpi</i>	1

- ▶ Strong adoption of base *R* packages (*parallel*)
  - ▶ Random numbers rarely handled properly
- ▶ MPI (a standard) not widely used
  - ▶ 3rd party dependency
  - ▶ Robust to user deployments
  - ▶ Error recovery
  - ▶ ...

# Algorithms

## Used

- ▶ Manager / worker
- ▶ lapply-like
- ▶ Interactive

*Ad hoc* user interactions

## Available

- ▶ pvec
- ▶ *snow*: subsets, sendData / recvData / ...
- ▶ *Rmpi*: rich MPI formulations
- ▶ Single instruction, multiple data (e.g., **pbdR**) and other models

# Pragmatic *Bioconductor* solutions

- ▶ Data structures
- ▶ Standard packaging
- ▶ Iteration
- ▶ The cloud



# Data structures

Use *de facto* standard data formats

- ▶ e.g., BAM, VCF files

Exploit column-oriented access

- ▶ e.g., *GRanges*-class: a single S4 instance
- ▶ More subtlety: *IRangesList*-class a single S4 instance with partitioning
- ▶ Key operations, e.g., `findOverlaps`, efficiently implemented

Exploit sparsity

- ▶ e.g., *Rle*-class: effectively compress whole-genome 'coverage'
- ▶ Supports rich set of operations

# Data structures

```
> gr
```

```
GRanges with 10 ranges and 2 metadata columns:
```

	seqnames	ranges	strand	score	GC
	<Rle>	<IRanges>	<Rle>	<integer>	<numeric>
a	chr1	[ 1, 10]	-	1	1
b	chr2	[ 2, 10]	+	2	0.88
c	chr2	[ 3, 10]	+	3	0.77
d	chr2	[ 4, 10]	*	4	0.66
e	chr1	[ 5, 10]	*	5	0.55
f	chr1	[ 6, 10]	+	6	0.44
g	chr3	[ 7, 10]	+	7	0.33
...					
---					

```
seqlengths:
```

chr1	chr2	chr3
1000	2000	1500

## Standard packaging: *BiocParallel*

Register parameterized back ends

- ▶ Sensible performance defaults
- ▶ Easy to switch between parallel & serial evaluation
- ▶ Scheduling of nested parallelism (to come)

Common signatures

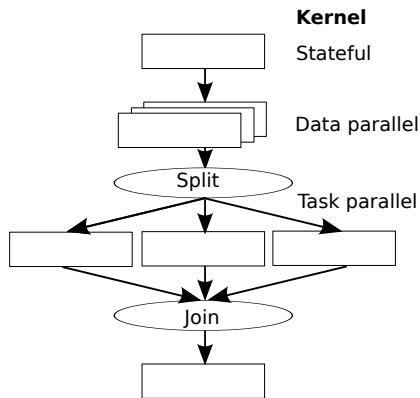
- ▶ `bplapply(X, FUN, ..., param)`, `bpvec(X, FUN, ..., param)`

Programming to contract, e.g., `bplapply`

- ▶ `X` must implement methods `length`, `[[`, and `[[`
- ▶ Currently: `mclapply` requires `as.list`, which defeats the purpose of some high-volume containers

## Iteration: *Streamer*

- ▶ *Producer* and *Consumer* kernels, assembled into streams
- ▶ yield output from a single chunk
- ▶ Requires on-line and other algorithms
- ▶ Formalism offers chance for code transformation / compilation



## Streamer

```
p <- Seq(to=50, yieldSize=5) # Producer: 1:50

param <- MulticoreParam(size=5)
team <- Team(function(x) {
  Sys.sleep(1); mean(x)
}, param=param)
s <- Stream(p, team)

system.time({
  while(length(y <- yield(s)))
    print(y)
}) ## about 2 seconds
```

# Streamer

```
dteam <-  
  DAGTeam(A=FunctionConsumer(function(y) y),  
          B=FunctionConsumer(function(A) -A),  
          C=FunctionConsumer(function(A) 1 / A),  
          D=FunctionConsumer(function(B, C) B + C))  
  
plot(dteam)  
  
strm <- Stream(Seq(to=10), dteam)  
sapply(strm, c)  
# [1]  0.00 -1.50 -2.67 -3.75 -4.80  
# [6] -5.83 -6.86 -7.88 -8.89 -9.90
```

# The cloud

- ▶ *Bioconductor* AMI, configured with, e.g., MPI support
- ▶ Helps address heterogeneity of user systems / administration
- ▶ Integration with *Galaxy* as a more 'user friendly' tool
- ▶ Unclear how this fits into academic / business funding models

# Recap

## *Bioconductor*

- ▶ Well-used
- ▶ Talented but not CS developers

Approaches to big data require. . .

- ▶ Efficient code, memory management, parallel evaluation

Pragmatic solutions

- ▶ Data structures, standard packaging, iteration, cloud



# Acknowledgements

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- ▶ NIH U41 HG004059